Detection of Herpes Simplex Viruses I andII in Cerebrospinal Fluid Specimens of Iraqi Children Presenting With Aseptic Meningitisby Using Real Time PCR Assay

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Abstract:

Introduction: Acute meningitis is a frequent syndrome encountered in emergency rooms. Viral meningitis is a common infectious disease of the central nervous system (CNS) that occurs worldwide. It can occur at any age but is most common in young children. Viral meningitis caused by herpes simplex virus (HSV) is a self-limited illness of 2–5 days' duration characterized by CSF findings of pleocytosis and an elevated protein level with a normal glucose level10. Although Herpes simplex I and II account for only 31% to 4%10f all cases of aseptic meningitis, and/or meningoencephalitis caused by HSV I and II are more important due to their potentialclinical severity in the case of meningoencephalit

Materials and Methods: CSF samples were collected from 68 patients suspected of having aseptic meningitis and/or encephalitis at Medical City Hospital, Baghdad, Iraq, during the period between April 2013 and June 2014. Viral DNA was isolated from 400 µl of a CSF specimen by using the high pure viral nucleic acid kit (Roche, USA) according to the manufacturer's instructions. DNA amplification was carried out on a Rotorgene 6000 (Corbett Life Sciences, Sydney, Australia) using Applied Biosystems (Roche, Branchburg, New Jersey) TaqMan Universal PCR Master Mix R.

Results: Twelve/68 (17.6%) CSF samples were positive for HSV-1whereas 2/68 (2.9%) samples were positive for HSV-2. There was no mixed infection. Herpesviruses were observed during the winter months. The positive samples were within age range 1-4 years. Of the 55 HSV DNA–positive cases, 5/12 (42%) occurred in male patients and 7/12 (58%) occurred in female patients. The same pattern was observed for negative samples, 23/56 (41%) male and 33/56 (59%) female.

Conclusion: HSV-1 and HSV-2 should be screened in cases of suspected aseptic meningitis. Further analysis of other agents and the stan \neg dardization of biology molecular methodologies are future proposition to improve the laboratorial diagnosis of viral meningitis in Iraq.

Keywords: Aseptic meningitis, HSV-1, HSV-2, children, Iraq, Real time PCR

I. Introduction

Acute meningitis is a frequent syndrome encountered in emergency rooms. Viruses and bacteria are the most common pathogens associated with this clinical picture¹.Viral meningitis is a common infectious disease of the central nervous system (CNS) that occursworldwide.It can occur at any age but is most common in young children².While the clinical course of viral meningitis and encephalitis may overlap, viral meningitis is usually self-limiting³, whereas the mortality from viral encephalitis ranges from 4.6% to 29%⁴⁻⁹.Viral meningitis caused by herpes simplex virus (HSV) is a self-limited illness of 2–5 days' duration characterized by CSF findings of pleocytosis and an elevated protein level with a normal glucose level¹⁰. Although Herpes simplex I and II account for only 31% to 4%¹ of all cases of aseptic meningitis, and/or meningoencephalitis caused by HSV I and II are more important due to their potential clinical severity in the case of meningoencephalitis¹¹.

Rapid etiologic diagnosis is very important to differentiate viral and bacterial meningitis so as to guide the introduction of antibiotic therapy early or avoid its unnecessary use for viral diseases.Conventional laboratory diagnostic methods, such as growth of a microorganism in culture and detection of specific antibody, are generally poor in the diagnosis of CNS infection. They are generally too slow, have low sensitivity and consequently they are of little use in diagnosing and treating the patient during the acute phase of the disease^{12,13}. Molecular techniques are now considered the gold standard for the detection in cerebrospinal fluid (CSF) samples of the viruses responsible for CNS infections^{14, 15}. Several studies have employed polymerase chain reaction (PCR) for diagnosing CNS infections, especially for HSV, studies have demonstrated that PCR is highly sensitive and specific, in addition to being fast and less invasive^{16, 17}.

There are few confirmed diagnoses of viral CNS infections in Iraq. Consequently, the true incidence and etiology of viral meningitis are not well described. The purpose of the present study was to screen for herpes simplex virus type 1 (HSV-1) and herpes simplex virus type 2 (HSV-2) in febrile hospitalized patients, utilizing molecular diagnostics to improve the laboratory diagnostic assessment of these patients.

II. Materials And Methods

Specimens CSF samples were collected from 68 patients suspected of having aseptic meningitis and/or encephalitis at Medical City Hospital, Baghdad, Iraq, during the period between April 2013 and June 2014. CSF samples were included in the study based on one or more signs of meningitis and/or encephalitis (severe headache, photophobia, nausea/vomiting, meningeal signs, petechial/purpural rash, altered mental status, seizures, and lethargy) and clinical indication for lumbar puncture, as determined by the attending physician.

CSF biochemical and cytological pattern of viral meningitis are: white blood cells>5 cells/mm³ with predominance of lymphocytes, CSF glucose normal (>45 mg/dL), CSF lactate <3.5 mmol/L,as well as the negative cultures for bacteria^{1, 18}.Patients' ages ranged from 1-4 years, 40 cases were male and 28cases were female. Upon receipt in the diagnostic laboratory, the specimens were processed or stored at - 20°C until the study.

Viral DNA extraction

Viral DNA was isolated from 400 μ l of a CSF specimen by using the high pure viral nucleic acid kit (Roche, USA) according to the manufacturer's instructions. DNA was stored at -20°C until PCR performed.

Real time PCR

PCR primers and probes were previously described¹⁰ (Table 1)

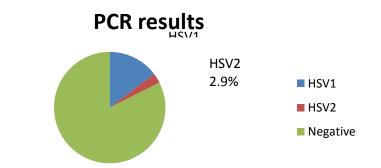
Table 1. P	Polymerase	Chain R	Reaction	Primers ar	nd Probes
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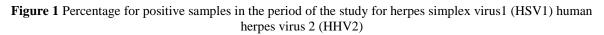
Virus (Target Gene)	Sense Primer 5 🗔	Antisense Primer 5 🗔 3 🗆	Probe 5 🗔 3 🗆
	3 🗆		
HSV-1 (gD)			
_	CGGCCGTGTGAC	CTCGTAAAATGGCCCCT	CCATACCGACCAC
	ACTATCG	CC	ACCGACGAACC
HSV-2 (gG)			
_	CGCTCTCGTAAAT	TCTACCCACAACAGAC	CGCGGAGACATTC
	GCTTCCCT	CCACG	GAGTACCAGATCG

DNA amplification was carried out on a Rotorgene 6000 (Corbett Life Sciences, Sydney, Australia) using Applied Biosystems (Roche, Branchburg, New Jersey) TaqMan Universal PCR Master Mix R. The PCR thermocycler conditions were 50°C for 2 minutes, 95°C for 10 minutes, 40 cycles of 95°C for 15 seconds, then 60°C for 1 minute and 60°C for 7 minutes¹⁰.

III. Results

A total of 68 CSF samples from patients with suspected aseptic meningitis were tested using the real time PCR protocol.Twelve/68 (17.6%) CSF samples were positive for HSV-1whereas 2/68 (2.9%) samples were positive for HSV-2 (figure 1). There was no mixed infection.





Herpesviruses were observed during the winter months. The positive samples were within age range 1-4 years. Of the 55 HSV DNA–positive cases, 5/12 (42%) occurred in male patients and 7/12 (58%) occurred in female patients. The same pattern was observed for negative samples, 23/56 (41%) male and 33/56 (59%) female.

IV. Discussion

Out of a total of 12/68 (17.6%) positive samples, 10 CSF samples were positive for HSV-1 and 2 CSF samples for HSV-2. Because of their neurotropic nature, HSV has been recognized as a relatively common CNS pathogen and the involvement of herpesviruses as etiologic agents of meningitis or encephalitis has been extensively described²⁰⁻²². When compared to other studies the percentage of herpes simplex virus positive specimens of our study is similar to that reported by other investigators such as 15% in Iran²³ while other investigators reported results differ from our results either lower such as 1.3% in Brazil²⁴ or higher such as 33% in Middle East region²⁵ these differences may be due different in sample criteria: Middle Eastern study is done on CSF specimens from encephalitis patients only and excluded meningitis patient and vice versa in Brazilian study while our study is done on CSF specimens from meningitis and/or encephalitis patients as well as our study included childhood only while Middle Easternand Brazilian studies are done on children and adults. Many studies also reported the positivity of HSV1 is higher than HSV2. The high number of CSF samples negative for HSV that suggests other viruses cause of aseptic meningitis such as enteroviruses, varicella zoster virus, adenovirus, measles, rubella and mumpsornon-infectious conditions including autoimmune diseases and carcinomatous meningitis must be considered²⁶. As well as different factors, including the time of sampling of the CSF specimens may be critical. Many studies observed that HSV DNA may be more readily detectable in the later phases after the onset of symptoms and not in the very acute phases^{27, 28}

The difficulty in making a laboratory diagnosis using traditional techniques in cases of aseptic meningitis and encephalitis has stimulated the development of studies using real time PCR assays.HSV- 1 and HSV-2 should be screened in cases of suspected aseptic meningitis. Further analysis of other agents and the standardization of biology molecular methodologies are future proposition to improve the laboratorial diagnosis of viral meningitis in Iraq.

References

- [1]. Kupila L, Vuorinen T, Vainionpaa R, Hukkanen V, Marttila RJ, Kotilainen P: Etiology of aseptic meningitis and encephalitis in an adult population. Neurology 2006, 66:75–80.
- [2]. Brown B, Oberste MS, Maher K, Pallansch MA. Complete genomic sequencing shows that polioviruses and members of human enterovirus species c are closely related in the noncapsid coding region. J Virology 2003; 77:8973-8984.
- [3]. Granerod J, Tam CC, Crowcroft NS, Davies NW, Borchert M, et al. (2010) Challenge of the unknown. A systematic review of acute encephalitis in nonoutbreak situations. Neurology 75: 924–932.
- [4]. Logan SA, MacMahon E (2008) Viral meningitis. BMJ 336: 36–40. Glaser CA, Honarmand S, Anderson LJ, Schnurr DP, Forghani B, et al. (2006) Beyond viruses: clinical profiles and etiologies associated with encephalitis. Clin Infect Dis 43: 1565–1577.
- [5]. Mailles A, Stahl JP (2009) Infectious encephalitis in france in 2007: a national prospective study. Clin Infect Dis 49: 1838–1847.
- [6]. Huppatz C, Durrheim DN, Levi C, Dalton C, Williams D, et al. (2009) Etiology of encephalitis in Australia, 1990–2007. Emerg Infect Dis 15: 1359–1365.
- [7]. Srey VH, Sadones H, Ong S, Mam M, Yim C, et al. (2002) Etiology of encephalitis syndrome among hospitalized children and adults in Takeo, Cambodia, 1999–2000. Am J Trop Med Hyg 66: 200–207.
- [8]. Granerod J, Ambrose HE, Davies NW, Clewley JP, Walsh AL, et al. (2010) Causes of encephalitis and differences in their clinical presentations in England: a multicentre, population-based prospective study. Lancet Infect Dis 10: 835–844.
- [9]. Le VT, Phan TQ, Do QH, Nguyen BH, Lam QB, et al. (2010) Viral etiology of encephalitis in children in southern Vietnam: results of a one-year prospective descriptive study. PLoSNegl Trop Dis 4: e854.
- [10]. Weidmann M, Meyer-Konig U, Hufert FT. Rapid detection of herpessimplex virus and varicella-zoster virus infections by real-time PCR.J ClinMicrobiol 2003; 41:1565–8.
- [11]. Sawyer MH: Enterovirus infections: diagnosis and treatment. CurrOpinPediatr 2001, 13:65-69.
- [12]. Lipton JD, Schafermeyer RW. Central nervous system infections: the usual and the unusual. Ped Emergencies 1995; 13:417-443.
- [13]. Anderson M. Management of cerebral infection. J NeurolNeurosurg Psychiatry 1993; 56:1243-1258.
- [14]. DeBiasi, R. L., and K. L. Tyler. 2004. Molecular methods for diagnosis of viral encephalitis. Clin. Microbiol. Rev. 17:903-925.
- [15]. Kimberlin, D. W. 2005. Diagnosis of herpes simplex virus infections of the CNS. Expert Rev. Mol. Diagn. 5:537–547.
- [16]. Guffond T, Dewilde A, Lobert P-E, Caparros-Lefebvre D, Hober D, Wattre P. Significance and clinical relevance of the detection of herpes simplex virus DNA by the polymerase chain in cerebrospinal fluid from patients with presumed encephalitis. Clin Infect Dis 1994; 18:744-49.
- [17]. Lakeman FD, Whitley RJ, and the National Institute of Allergy and Infectious Diseases Collaborative Antiviral Study Group. Diagnosis of herpes simplex encephalitis: application of polymerase chain reaction to cerebrospinal fluid from brain-biopsied patients and correlation with disease. J Infect Dis 1995; 171:857-863.
- [18]. Rice P. (2005). Viral meningitis and encephalitis. Medicine. 33, 60-63.
- [19]. Azadfar S., Cheraghali F., Moradi A., Javid N., Tabarraei A. Herpes Simplex Virus Meningitis in Children in South East of Caspian Sea, Iran. Jundishapur J Microbiol. 2014; 7: e8599
- [20]. Frantzidou F, Kamaria F, Dumaidi K, Skoura I, Antoniadis A, Papa A. Aseptic meningitis and encephalitis because of herpesviruses and enteroviruses in an immunocomptent adult population. Eur J Neurol 2008; 15:995-997.
- [21]. Gilden DH, Mahalingam R, Cohrs RJ, Tyler KL. Herpesvirus infections of the nervous system. Nat ClinPract 2007; 3:82-94.

- [22]. Berger JK, Houff S. Neurological complications of herpes simplex virus type 2 infection. Clinical implications of basic neuroscience research 2008; 63: 396-399.
- [23]. Ghannad M., Solgi G., Hashemi S., Zebarjady-Bagherpour J., Hemmatzadeh A., Hajilooi M. Herpes Simplex Virus Encephalitis in Hamadan, Iran.IRAN. J. MICROBIOL. Vol. 5, No. 3 (2013), 272-277.
- [24]. Vida L., Almeida S., Messias-Reason I., Nogueira M., Debur M., Pessa L., Pereira L., Rotta I, Takahashi G., Silveira C., Araújo J., Raboni S. Enterovirus and herpesviridae family as etiologic agents of lymphomonocytary meningitis, Southern Brazil. ArqNeuropsiquiatr 2011;69(3):475-481.
- [25]. Ibrahim A.,Obeid M., JoumaM.,Roemer K.,Mueller-LantzschN.,and Gartner B. Prevalence of Herpes Simplex Virus (Types 1 and 2), Varicella-Zoster Virus, Cytomegalovirus, and Human Herpesvirus 6 and 7 DNA in Cerebrospinal Fluid of Middle Eastern Patients with Encephalitis. J. CLIN. MICROBIOL. Vol. 43, No. 8 (2005), p. 4172–4174.
- [26]. Almeida S M, Nogueira M B, Raboni S M, Vidal L R R. Laboratorial diagnosis of lymphocytic meningitis. Brazilian J Infect Dis 2007; 11:478-484.
- [27]. Aurelius, E., B. Johansson, B. Skoldenberg, A. Staland, and M. Forsgren. 1991. Rapid diagnosis of herpes simplex encephalitis by nested polymerase chain reaction assay of cerebrospinal fluid. Lancet 337:189–192.
- [28]. Davies, N. W., L. J. Brown, J. Gonde, D. Irish, R. O. Robinson, A. V. Swan, J. Banatvala, R. S. Howard, M. K. Sharief, and P. Muir. 2005. Factors influencing PCR detection of viruses in cerebrospinal fluid of patients with suspected CNS infections. J. Neurol. Neurosurg. Psychiatry 76:82–87.